

IN THE SPECIFICATION:

Please amend the specification as follows.

At page 13, lines 10-20, please amend the second full paragraph on that page as follows:

Figure 20 demonstrates migration of HUVECs in wound assay. The wounded monolayer of HUVEC was exposed for 16 hours to respective aliquots of conditioned media. Migrated cells were then fixed, stained and photographed. (A) Representative micrographs of migrated HUVECs in WT-hPTTG-CM are shown. 1a, conditioned media alone; 2b, conditioned media + 100 ng/ml anti-bFGF antibody. (B) Quantification of migrated HUVECs. The number of cells within 0.1 x 2.5-mm area in three fields was counted using the original mark made by razor blade as origin. The results shown are the average number of cells \pm SD per field of three separate experiments. 1a, control; 2b, WT-hPTTG-CM; 3e, M-hPTTG-CM; 4d, C-CM; 5e, N-CM; open circle, 1 ng/ml bFGF in DMEM; open triangle, serum-free DMEM; closed circle, conditioned medium alone; closed triangle, conditioned medium + 100 ng/ml anti-bFGF antibody; closed square, conditioned medium + 100 ng/ml pre-immune goat IgG.

At page 13, lines 21-28, please amend the third full paragraph on that page as follows:

Figure 21A shows migration of HUVECs in a modified Boyden chamber assay. The sample conditioned medium was placed in the lower chamber and HUVECs were added in the upper chamber of a modified Boyden chamber. After 24 hours incubation, non-migrating cells were removed and cells migrated through membrane pores (8 μ m) were stained with Giemsa and photographed. (A)-Micrographs of migrated HUVECs from representative membranes under each experimental condition. 1a, serum-free DMEM; 2b, 1 ng/ml bFGF in DMEM; 3e, WT-hPTTG-CM; 4d, WT-hPTTG-CM + 100 ng/ml anti-bFGF antibody; 5e, WT-hPTTG-CM + 100 ng/ml pre-immune goat IgG; 6f, M-hPTTG-CM; 7g, C-CM; 8h, N-CM.

At page 14, lines 4-15, please amend the first full paragraph on that page as follows:

Figure 22 demonstrates tube-formation of HUVECs on Matrigel. 5×10^4 of HUVECs suspended in sample conditioned media and plated on GFR Matrigel thickly coated 24-well culture plates. After 24 hours incubation, cells were photographed under phase-contrast microscopy. Figure 22A shows micrographs of tube-forming HUVECs. Representative photographs for each experimental condition are shown. 1a, serum-free DMEM; 2b, 1 ng/ml bFGF in DMEM; 3e, WT-hPTTG-CM; 4d, WT-hPTTG-CM + 100 ng/ml anti-bFGF antibody; 5e, WT-hPTTG-CM + 100 ng/ml pre-immune goat IgG; 6f, M-hPTTG-CM; 7g, C-CM; 8h, N-CM. Figure 22B shows quantification of tube-formation. Tube length was quantified as described in "Materials and Methods". The mean pixel number \pm SD of three separate experiments is expressed. From left to right: D, serum-free DMEM; F, 1 ng/ml bFGF in DMEM; WT, WT-hPTTG-CM; M, M-hPTTG-CM; C, C-CM; N, N-CM. $p < 0.01$ versus * DMEM, ** WT-hPTTG-CM, *** respective conditioned medium (CM) alone.

At page 14, lines 16-24, please amend the second full paragraph on that page as follows:

Figure 23 demonstrates vascular reactions of CAM to conditioned media. Test samples, positive or negative control in collagen sponges were loaded on CAM of 9-day-old chick embryos. After 4 days, CAMs were photographed. (A) photographs of representative CAMs of 13-day-old chick embryo. 1a, serum-free DMEM; 2b, 1 ng/ml bFGF in DMEM; 3e, WT-hPTTG-CM; 4d, M-hPTTG-CM; 5e, C-CM; 6f, N-CM. (B) Quantification of induced vessels. Number of blood vessels entering the collagen sponges was counted under stereomicroscopy. The data shown is the mean \pm SD of three separate experiments. D, serum-free DMEM; F, 1 ng/ml of bFGF in PBS; WT, WT-hPTTG-CM; M, M-hPTTG-CM; C, C-CM; N, N-CM. $p < 0.01$ versus * PBS, ** WT- hPTTG-CM.